

Replace the second full paragraph on page 4 (lines 17-24) with the following new paragraph rewritten in clean form:

Sub B2
A2

~~Any of the fibronectin type III domain-containing proteins described herein~~ may be formulated as part of a fusion protein (for example, a fusion protein which further includes an immunoglobulin F_c domain, a complement protein, a toxin protein, or an albumin protein). In addition, any of the fibronectin type III domain proteins may be covalently bound to a nucleic acid (for example, an RNA), and the nucleic acid may encode the protein. Moreover, the protein may be a multimer, or, particularly if it lacks an integrin-binding motif, it may be ~~formulated in a physiologically-acceptable carrier.~~

Replace the final partial paragraph on page 4 (line 25) with the following new partial paragraph rewritten in clean form:

N.E. The present invention also features proteins that include a

Replace the fourth paragraph on page 11 (lines 10-14) with the following new paragraph rewritten in clean form:

Sub B4
A3

~~FIGURE 5 is a photograph showing the structural similarities between a~~ ¹⁰F_n3 domain and 15 related proteins, including fibronectins, tenascins, collagens, and undulin. In this photograph, the regions are labeled as follows: constant, dark blue; conserved, light blue; neutral, white; variable, red; and RGD integrin-binding motif (variable), ~~yellow.~~

Replace the fifth paragraph on page 11 (lines 15-19) with the following new paragraph rewritten in clean form:

Sub B5
A4

~~FIGURE 6 is a photograph showing space filling models of fibronectin III~~ modules 9 and 10, in each of two different orientations. The two modules and the integrin binding-loop (RGD) are labeled. In this figure, blue indicates positively charged residues, red indicates negatively charged residues, and white indicates

A4

~~uncharged residues~~

Replace the fourth paragraph on page 13 (lines 20-24) with the following new paragraph rewritten in clean form:

~~The antibody mimics of the present invention are based on the structure of a fibronectin module of type III (Fn3), a common domain found in mammalian blood and structural proteins. This domain occurs more than 400 times in the protein sequence database and has been estimated to occur in 2% of the proteins sequenced to date, including fibronectins, tenascin, intracellular cytoskeletal~~

Replace the second partial paragraph on page 19 (lines 14-24) with the following new partial paragraph rewritten in clean form:

~~The antibody mimics described herein may be used in any technique for evolving new or improved binding proteins. In one particular example, the target of binding is immobilized on a solid support, such as a column resin or microtiter plate well, and the target contacted with a library of candidate scaffold-based binding proteins. Such a library may consist of ¹⁰F_n3 clones constructed from the wild type ¹⁰F_n3 scaffold through randomization of the sequence and/or the length of the ¹⁰F_n3 CDR-like loops. If desired, this library may be an RNA-protein fusion library generated, for example, by the techniques described in Szostak et al., U.S.S.N. 09/007,005, now U.S. Patent No. 6,258,558 B1, and 09/247,190, now U.S. Patent No. 6,261,804 B1; Szostak et al., WO98/31700; and Roberts & Szostak, Proc. Natl. Acad. Sci. USA (1997) vol. 94, p. 12297-12302. Alternatively, it may be a DNA-protein library (for example, as described in~~

Replace the first partial paragraph on page 20 (lines 1-9) with the following new partial paragraph rewritten in clean form:

~~Lohse, DNA-Protein Fusions and Uses Thereof, U.S.S.N. 60/110,549, filed December 2, 1998 and 09/453,190, filed December 2, 1999). The fusion library is incubated with the immobilized target, the support is washed to remove non-specific binders, and the tightest binders are eluted under very stringent~~

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~~conditions and subjected to PCR to recover the sequence information or to create a new library of binders which may be used to repeat the selection process, with or without further mutagenesis of the sequence. A number of rounds of selection may be performed until binders of sufficient affinity for the antigen are obtained.~~

Replace the last paragraph on page 24 (lines 18-20) with the following new paragraph rewritten in clean form:

usab
A8

~~Unispl-s (splint oligonucleotide used to ligate mRNA to the puromycin-containing linker, described by Roberts et al, 1997, supra): 5'-TTTTTTTTTINAGCGGATGC-3' (SEQ ID NO: 13)~~

Replace the second paragraph on page 25 (lines 10-22) with the following new paragraph rewritten in clean form:

usab
A9

~~Next, each of the double stranded fragments was transformed into a RNA-protein fusion (PROfusion™) using the technique developed by Szostak et al., U.S.S.N. 09/007,005, now U.S. Patent No. 6,258,558 B1, and U.S.S.N. 09/247,190, now U.S. Patent No. 6,261,804 B1; Szostak et al., WO98/31700; and Roberts & Szostak, Proc. Natl. Acad. Sci. USA (1997) vol. 94, p. 12297-12302. Briefly, the fragments were transcribed using an Ambion *in vitro* transcription kit, MEGAshortscript (Ambion, Austin, TX), and the resulting mRNA was gel-purified and ligated to a DNA-puromycin linker using DNA ligase. The mRNA-DNA-puromycin molecule was then translated using the Ambion rabbit reticulocyte lysate-based translation kit. The resulting mRNA-DNA-puromycin-protein PROfusion™ was purified using Oligo(dT) cellulose, and a complementary DNA strand was synthesized using reverse transcriptase and the RT primers described above (Unisplint-S or flagASA), following the manufacturer's instructions.~~

Replace the first partial paragraph on page 27 (lines 1-3) with the following new partial paragraph rewritten in clean form:

N.E. from the master library following the general procedure described in Szostak et al.,